## **Supplementary Information**

**Title:** Tetracycline-inactivating enzymes from environmental, human commensal, and pathogenic bacteria cause broad-spectrum tetracycline resistance

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## **Supplementary Figures**



Supplementary Figure 1: Tet(X7) inactivates latest-generation tetracyclines by monooxygenation. (a) *In vitro* reactions containing purified Tet(X7), NADPH, and tigecycline, omadacycline, or eravacycline were assayed by absorbance scans taken at one minute intervals. The rainbow pattern depicts a spectral change over time. Decrease in absorbance over time from 360 nm to 400 nm indicates enzymatic disruption of the characteristic tetracycline  $\beta$ -diketone chromophore and consumption of NADPH. (b) In the case of tigecycline, omadacycline, and eravacycline, the product of enzymatic modification is consistent with monooxygenation. Plots show the relative extracted ion counts of the parent tetracycline (blue curves; *m/z* for [M+H]<sup>+</sup> equal to 586, 557, 559, respectively ) or the monooxygenated product (orange curves; *m/z* for [M+H]<sup>+</sup> equal to 602, 573, 575, respectively). (c) HPLC analysis with optical absorbance at 260 nm confirms enzyme-dependent degradation of tigecycline, eravacycline, and omadacycline by Tet(X7).



Supplementary Figure 2: Product of Tet(X7) hydroxylation of tigecycline coelutes with product of Tet(X) hydroxylation of tigecycline. *In vitro* reactions containing NADPH, tigecycline, and purified Tet(X) (a, b) or Tet(X7) (c, d) assayed by LC-MS indicate conversion of tigecycline (m/z for  $[M+H]^+$  equal to 586) to monooxygenated tigecycline (m/z for  $[M+H]^+$  equal to 602) over 120 minutes. Plots show extracted ion count LC-MS chromatograms for tigecycline (e) and monooxygenated tigecycline (f) from reactions with Tet(X) and Tet(X7) after 120 minutes. Signals were smoothed in ChemStation software using the "smooth all signals" function with a Gaussian fit of 0.05 min and a step size of 0.156 min. Also included are no enzyme controls and equimolar mixtures of the products of reactions with both enzymes. The products of Tet(X) and Tet(X7) monooxygenation of tigecycline coelute.



Supplementary Figure 3: Structural comparison of Tet(X7) and Tet(X) structures.

Top view: Two proteins are shown in cartoon (left: Tet(X7) and right: Tet(X)). Bound FAD is shown in grey (C-grey, O-red, N-blue) spheres in both the structures. The bound tigecycline in Tet(X) (PDB ID: 4A6N) is shown in green (C-green, O-red, N-blue) spheres. The conserved residues between two proteins are shown as lines and all different residues are shown in stick representation. In Tet(X7) (left), the divergent residues are colored as blue and in Tet(X), the divergent residues were colored based on the comparative physiological properties of residues in the two proteins. Blue represents the most similar substitution and the red represents the most contrasting substitution. Bottom view: The y-axis is rotated by 90° (counterclockwise) and the bridge-helix is shown in zoom.

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u	1	tet(X)	M	RIDTDKQMNI	LL <mark>SDKN</mark> VAII	GGGPVGL <mark></mark> MA	<u>KLLQQNGIDVSVYERDND</u> R	EARIFGGTL
	2	tet(X3)	M M	RIDTDKOMNI	LL <mark>SDKN</mark> VAII	GGGPVGL <mark>O</mark> MA	<u>KLLQQNGIDVSVYERDND</u> R	EARIFGGTL
	3	tet(X4)	MS-	NKEKOMNI	LL <mark>SDKN</mark> VAII	GGGPVGL <mark>O</mark> MA	<b>KLLQQNGIDVSVYERDND</b> R	EARIFGGTL
	4	tet(X7)		MTI	LKN <mark>K</mark> KITII	GAGPVGL <mark>O</mark> MA	RLLOONRVDVIIVYERDKDR	DARIFGGTL
					_			
			61	•	•	•	1.	. 120
	1	tet(X)	DLH	K <mark>GSGQE</mark> AMKK	AGLLQTYYD	LALPMGV <mark>N</mark> IA	DKKGNILSTKNVKPENRFD	NP <mark>EINRND</mark> L
	2	tet(X3)	DLH	K <mark>GSGQE</mark> AM <mark>K</mark> K	AGLLQTYYD	LALPMGV <mark>N</mark> IA	D <mark>EKGNILSTKNVKP</mark> ENRFD	NP <mark>EINRND</mark> L
	3	tet(X4)	DLH	KG <mark>SGQE</mark> AM <mark>K</mark> K	AGLLOTYYD	LALPMGV <mark>N</mark> IA	D <mark>E</mark> KGNILSTKNVKPENRFD	NPEINRNDL
	4	tet(X7)	DLH	RD <mark>SGQE</mark> AM <mark>K</mark> F	<b>AGLLQTYYD</b>	LALPMGV <mark>N</mark> IA	D <mark>EKGNILTTKNVKPENRFD</mark>	NP <mark>EINRND</mark> L
			121	•	•	:		. 180
	1	tet(X)	RAI	LLNSLENDTV	/IW <mark>DRK</mark> LVML	EPG <mark>KKK</mark> WTLT	F <mark>ENK</mark> PSET <mark>AD</mark> LVILANGGM	S <mark>KVRKFVTD</mark>
	2	tet(X3)	RAI	LLNSLENDTV	/IW <mark>DRK</mark> LVML	EPG <mark>KKK</mark> WTLT	FENK <mark>PSET</mark> ADLVILANGGM	SKIRSFVTD
	3	tet(X4)	RAI	LLNSLENDTV	/IW <mark>DRK</mark> LVML	EPG <mark>KKK</mark> WTLT	FENK <mark>P SET</mark> ADLVI I <mark>AN</mark> GGM	S <mark>KVRKFVTD</mark>
	4	tet(X7)	RTI	LLNSLQNDT	/IW <mark>DRK</mark> LV <mark>T</mark> L	EPD <mark>K</mark> EKWTLT	FE <mark>DK</mark> PSETADLVI IANGGM	<mark>S</mark> KVRKFVTD
			181	•	2	•	• •	. 240
	1	tet(X)	TEV	EETGTFN I QA	A <mark>DIHQPEIN</mark> C	PGFFQL <mark>C</mark> NGN	RLMASHQGNLLFANPNNNG	ALHFGI <mark>s</mark> fk
	2	tet(X3)	<mark>T</mark> Q <b>V</b>	EETGTFNIQ#	A <mark>DIIQPEINC</mark>	PGFFQ <mark>LC</mark> NGN	<mark>RLMA</mark> GHQGILLFAN <mark>PNNN</mark> G	ALYL <mark>GI S</mark> F <mark>K</mark>
	3	tet(X4)	TEV	EETGTFNIQ#	DIHHPEVNC	PGFFQL <mark>C</mark> NGN	RLMAAHQGNLLFANPNNNG	ALHFGI <mark>S</mark> FK
	4	tet(X7)	TEV	EETGTFNIQ#	DIHQPEUNC	PGFFQL <mark>C</mark> NGN	RLMAAHQGNLLFANPNNNG	ALHFGI <mark>S</mark> F <mark>K</mark>
			241	:	•	•		3 300
	1	tet(X)	TPD	EWKNQTQVDF	<u>'QNRNSVVD</u> F	LL <mark>KEFSDWDE</mark>	RYKELIHTTLSFVGLATRI	FPL <mark>EK</mark> PWKS
	2	tet(X3)	TPD	EWKNKIPLDF	rq <mark>drnsva</mark> df	LL <mark>K</mark> RF <mark>S</mark> KWSE	VYK <mark>QLI</mark> RSVSTFQCLPTRK	FPLNNDWKS
	3	tet(X4)	TPD	EWKNQTQVDF	FQNRNSVVDF	LL <mark>KE</mark> FSDWDE	RYKELIRVTSSFVGLATRI	FPLG <mark>K</mark> SWKS
	4	tet(X7)	<b>T</b> SD	EWKSKTLVDF	<sup>r</sup> Qd <mark>rnsvvd</mark> f	LL <mark>K</mark> K <mark>FSDWDE</mark>	RYKELIRVTSSFVGLATRI	FPLC <mark>K</mark> SWKS
			301	•	•	•	· ·	. 360
	1	tet(X)	KRP	LPI MIGDAA	AHLMPPFAGQ	GVNSGLVDAL	ILSDNLADGKENS I DEAVE	NYEQQMFIY
	2	tet(X3)	NRP	LPIIMIGDAA	AHLMSPFAGQ	GVNTGLLDAL	ILSENLTNGEFTS IENAIE	NYEQQMFVY
	3	tet(X4)	KRP		AHLMPPFAGO	GVNSGLMDAL	ILSONLTNG <mark>KENS I DE</mark> AIE	NYEQQMFTY
	4	tet(X/)	KRP	LPI <mark>O</mark> MIG <mark>D</mark> AA	LMPPFAG	GV <mark>NS</mark> GLM <mark>D</mark> AL	ILSONLTNG <mark>KFNSIEE</mark> AIE	NYEQQMFAY
			261					
	1	$\pm a \pm (N)$	361			J 388		
	1	tet(X)	GKD					
	2	tet(X3)		IQDESTENET				
	כ ⊿	tet(X4)						
4 Tet (X/) GREADANSIIIMETEMESSDESECKIMAL								
h	Г		$tot(\mathbf{V})$	tot(XA)	$tot(\sqrt{7})$	tot(V2)		
Ŋ	Ļ	( ()))						
		tet(X)	100	95.06	85.71	85.57		
	Ī	tet(X4)	95.06	100	88.89	85.45		
	ľ	tet(X7)	85 71	88.89	100	80.95		
	F	tot(V2)	05 57	05 15	90 0F	100		
			00.07	00.40	00.90			

**Supplementary Figure 4: Sequence comparison of clinically implicated** *tet(X)* **variants.** Multiple sequence alignment (A) and percent identity matrix (B) of clinically implicated *tet*(X) variants. Residues in multiple sequence alignment are colored when identical to *tet*(X) based on amino acid property according to default MView color palette<sup>63</sup>.

Primer sequence $(5' \rightarrow 3')$	Designed specificity
ΑΤGACTTTGCTAAAAAATAAAAAAATA	TE_7F_Contig_3 (452_1588)
TTATAGATTCATTAGTTTTTGGAATGA	TE_7F_Contig_3 (452_1588)
ATGAATTTACTAAACAATAAAAAAGTTACA	TG_0402_Contig_27 (425_1561)
TTATAGATTCATTAGTTTTTGGAATGA	TG_0402_Contig_27 (425_1561)
ATGACAATGCGAATAGATACAGACA	TE_0402_Contig_1789 (2156_3322)
TTATACATTTAACAATTGCTGAAACG	TE_0402_Contig_1789 (2156_3322)
ATGACTTTGCTAAAAAATAAAAAAATT	TE_0402_Contig_1211 (2189_3325)
TTATAGATTCATTAGTTTTTGGAATGA	TE_0402_Contig_1211 (2189_3325)
ATGACAATGCGAATAGATACAGACA	TG_0402_Contig_45 (861_2027)
TTATACATTTAACAATTGCTGAAACG	TG_0402_Contig_45 (861_2027)
ATGACAATGCGAATAGATACAGACA	TG_0401_3a_Contig_236 (181_1347)
TTATACATTTAACAATTGCTGAAACG	TG_0401_3a_Contig_236 (181_1347)
ΑΤGACTTTACTAAAACATAAAAAAATTACA	TE_6F_Contig_7 (1101_2237)
TTATAGATTCATTAGTTTTTGGAATGA	TE_6F_Contig_7 (1101_2237)
ATGACTAGCGATAAGAACCCT	S08_TE.2:1046-1630 Contig:2
TCATTCGTATTCTGGTAGCG	S08_TE.2:1046-1630 Contig:2

Supplementary Table 1 Primers used in this study